

Dealing with "some" important statistical assumptions.

1) The issue of normality (today):

- Parametric (e.g., ANOVA): assume parametrized families of probability distributions (e.g., normal defined by two parameters, i.e., mean and variance). Parameter estimates tend to be sensitive to nonnormality (e.g., issue in regression slopes), but not necessarily in statistical hypothesis testing (P-values may be not as sensitive).

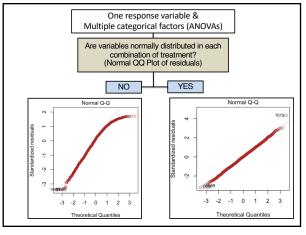
- Non-parametric: either distribution free (e.g., permutation tests) or ranked based tests.

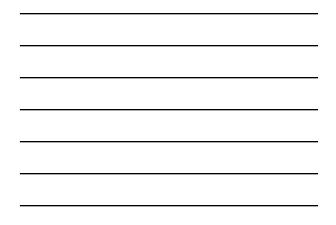
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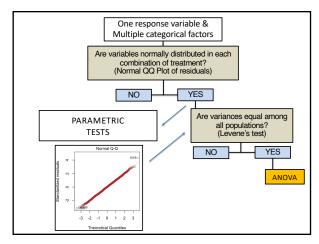
Dealing with "some" important statistical assumptions.

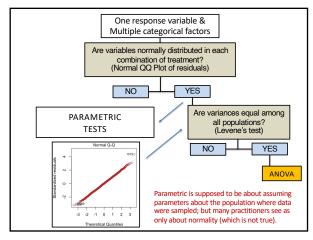
2) The issue of homogeneity of variances (later in the course):

- Standard (e.g., ANOVAs, regressions) assume homoscedasticity.
- Robust approaches (Welch's ANOVA, Weighted least squares) are good to deal with heteroscedasticity.

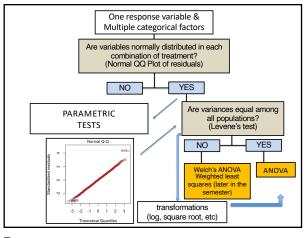




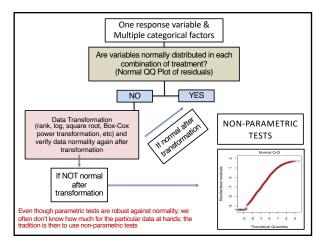




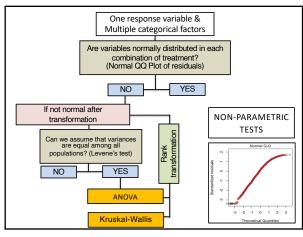




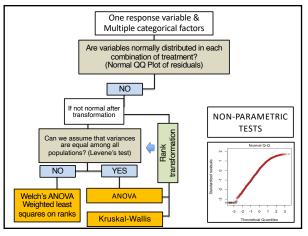


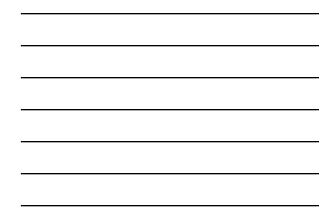


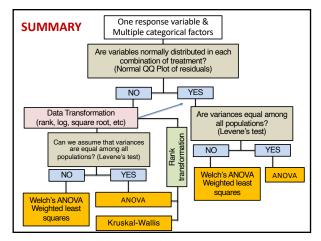




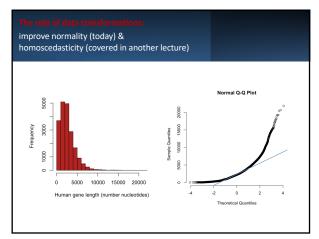




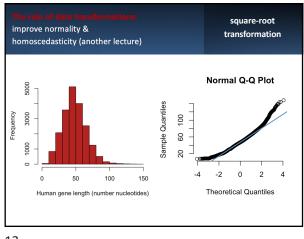




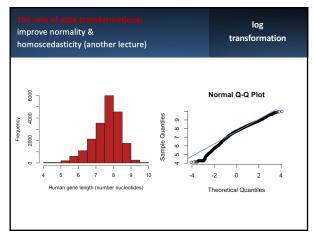














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A few words on data transformation

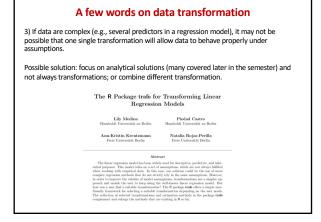
One size may not fit all:

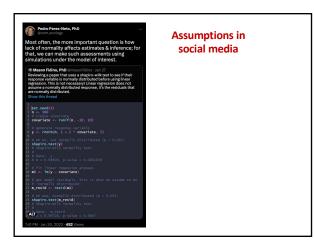
1) One transformation may help approximate normality, but another transformation may be required to approximate homoscedasticity (e.g., log(sqrt(data))).

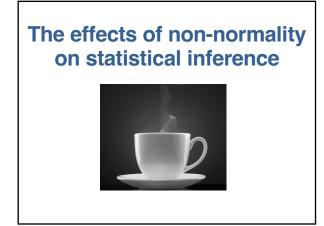
2) One transformation may negate (reverse) the other – the one that makes the data approximate homoscedasticity may make data non-longer normal.

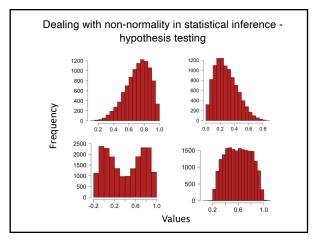
3) If data are complex (e.g., several predictors in a regression model), it may not be possible that one single transformation will allow data to behave properly under assumptions.

Possible solution: focus on analytical solutions (many covered in this course) and not always transformations; or combine different transformation.

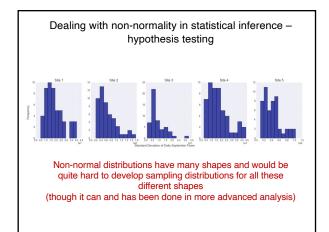












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The effects of non-normality on statistical test

Parametric tests assuming normality (e.g., t-test & ANOVA) are often robust against non-normality; but depending on the type of non-normality (shape), parametric tests can have type I errors different (often greater) from alpha; and low power (increased type II error).

One challenge is to separate normality from heteroscedasticity issues (even in simulations).

The other challenge is when samples come all from populations with different distributions (even though they could have the same means, i.e., $H_{\rm 0}$ is true).

The effects of non-normality on statistical test

Parametric tests assuming normality (e.g., t-test & ANOVA) are often robust against non-normality; but depending on the type of nonnormality (shape of the distribution), parametric tests can have type I errors (false positives) that differ (often greater) from alpha; and low power (increased type II error; false negatives).

Br J Math Stat Psychol. 2013 May.86(2):224-44. doi: 10.1111/j.2044-8317.2012.02047.x. Epub 2012 May 24. The impact of sample non-normality on ANOVA and alternative methods.

Lantz B¹. Author information

With the information
Abstract
In provide the information
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In provide the information of normality is doubtiful. The conceptual problem with this approach is that such a two-stage process makes both the power and the significance of the entire procedure uncertain, as type I and type II encore are possible at both stages, the sample with both approach is the significance of the entire procedure uncertain, as type I and type II encore are possible at both stages, the sample with bothoush process the proceeding to the dross of the significance of the entire procedure uncertain, as type I and type II encore are possible at both stages.
A type I encore the stages, for example, with bothoush process the proceeding of a public theory approach appendix the power of the significance of the entire procedure uncertain.
Schmidter et al. (2010), which proposes that simulated sets of sample data bar ranked with negrect to ther degree of normality.
Brown-Forsynte test, Weich test, and Kruskal-Weils test when used with different distributions, sample romain with weight to proceed on explaining and effect sizes. The overall and ethough therefore to be projectioned on the strange formation of the strange formation of the source procession and effect sizes. The overall and ethough therefore to be produced on explaining and distribution are not at least

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What happens if the Type I error probability (rate) is greater than alpha? i.e., increase number of False Positives.

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What happens if the Type I error probability (rate) is greater than alpha? i.e., increase number of False Positives.

What happens if the Type I error probability (rate) is smaller than alpha? decrease False Positives but also decrease True Positives (i.e., lower statistical power).

Type I versus Type II errors – the "common" view

A **Type I error (false positive)** is an **error** in every sense of the word. A conclusion is drawn that the null hypothesis is false when, in fact, it is true.

Therefore, **Type I** errors are generally considered more serious than **Type II** errors (false negatives).

Type II errors are often considered as "oh well, we were not able to detect an effect"...perhaps increase sample size!

Adapted from http://davidmlane.com/hyperstat/A2917.html

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When committing a type I error, you are stating that something that is false to be true.

CONFUSING: When committing a type II error, you are NOT stating that something that is true to be false (you are just not discovering something new).

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Non-parametric tests based on ranks are those that can handle non-normal data

These are the main tests traditionally used in Biology for comparing samples:

1) For comparing two samples (analogue of the parametric two sample t-test) – *The Mann–Whitney U-test* (also known as the Mann–Whitney–Wilcoxon test, the Wilcoxon rank-sum test, or the Wilcoxon two-sample test).

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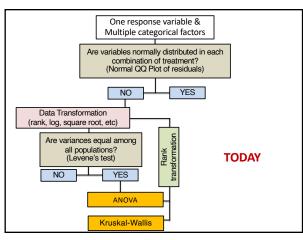
1) For comparing two samples (analogue of the parametric two sample t-test) – *The Mann–Whitney U-test* (also known as the Mann–Whitney–Wilcoxon test, the Wilcoxon rank-sum test, or the Wilcoxon two-sample test).

2) For comparing multiple samples (analogue of the parametric ANOVA) – *The Kruskal-Wallis test* (generalization of the U-test)

The P-value for the *The Mann–Whitney U-test and the The Kruskal-Wallis test* is mathematically the same; as such, we will cover only the latter.

Note: remember that $t^2 = F$; we often cover t-tests (and not only ANOVAs) in courses for two main reasons – [1] one sample t-tests; [2] understand the nature of post-hoc testing (e.g., post-hoc pairwise comparisons of means after ANOVA and because there is a t-test dealing with samples when their populations differ in their variances).

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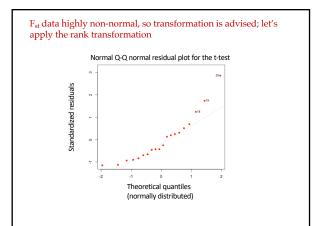


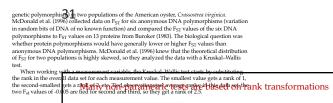
genetic polymorphism) in two populations of the American oyster, Crassestra virginica. McDonald et al. (1996) collected data on Fg. for six anonymous DNA polymorphisms (variation in random bits of DNA on oh known function) and compared the Fg. values of the six DNA polymorphisms to Fg. values on 13 proteins from Buroker (1983). The biological question was whether protein polymorphisms would have generally lower or higher Fg. values than anonymous DNA polymorphisms. McDonald et al. (1996) have that the theoretical distribution of Fg. for two populations is highly skewed, so they analyzed the data with a Kruskal-Wallis test. When working with a measurement variable, the Kruskal-Wallis tod states?

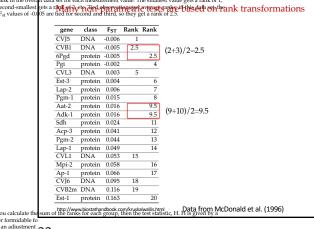
gene	class	FST	Example: Fst is a measure of the amount of
CVJ5	DNA	-0.006	geographic variation in a genetic polymorphisr Here, McDonald et al. (1996) compared two populations of the American oyster regarding t based on six anonymous DNA polymorphisms (variation in random bits of DNA of no known
CVB1	DNA	-0.005	
6Pgd	protein	-0.005	
Pgi	protein	-0.002	
CVL3	DNA	0.003	
Est-3	protein	0.004	
Lap-2	protein	0.006	function) and compared them to Fst values on proteins.
Pgm-1	protein	0.015	
Aat-2	protein	0.016	
Adk-1	protein	0.016	Question: Do protein differ in Fst values in con to anonymous DNA polymorphisms?
Sdh	protein	0.024	
Acp-3	protein	tein 0.041	, I , I
Pgm-2	protein	0.044	Zero Fst = no genetic variation (panmictic)
Lap-1	protein	0.049	negative Fst = more genetic variation within
CVL1	DNA	0.053	populations than between the two populations bei compared. positive Fsr = more variation between populations within the two populations being compared.
Mpi-2	protein	0.058	
Ap-1	protein	0.066	
CVJ6	DNA	0.095	
CVB2m	DNA	0.116	
Est-1	protein	0.163	

that the probability of the P value correspon groups minus 1. For **t30** protein is 10.68, H=0.0 hypothesis that the F₅ rejected.



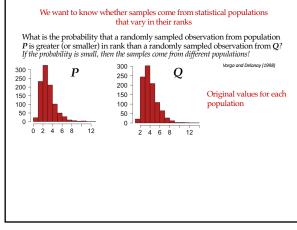




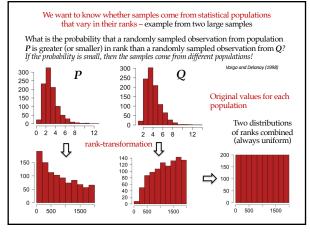


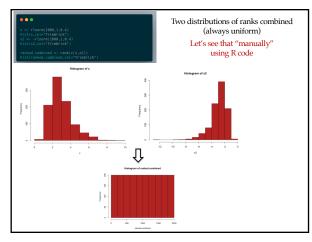
You calculate the sum of rather formidable fo with an adjustment that the probability '32 the P value correspond groups minus 1. For protein is 10.68, H=(hypothesis that the 1 rejected

For the reasons given above, I think it would actually be better to analyze the oyster data with one-way anova. It gives a P value of 0.75, which fortunately would not change the conclusions of McDonald et al.

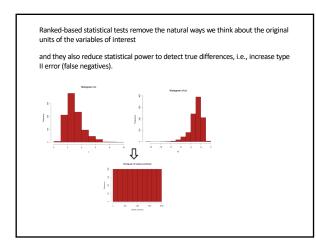










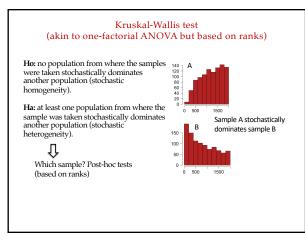












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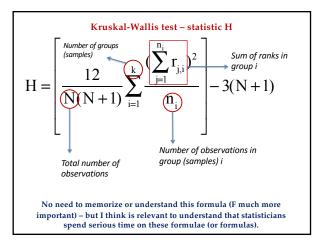
Kruskal-Wallis test (akin to one-factorial ANOVA but based on ranks)

 ${\bf H}\!{\bf c}$ no population from where the samples were taken stochastically dominates another population (stochastic homogeneity).

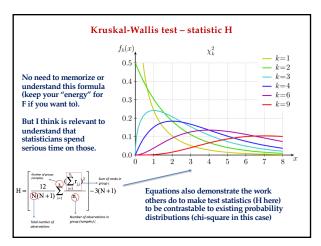
Ha: at least one population from where the sample was taken stochastically dominates another population (stochastic heterogeneity).

FSTs data __________ Ho: DNA and protein do not stochastically dominate each other in their FSTs.

 \mathbf{H}_{A} : Either DNA or protein stochastically dominate each other in their $\mathrm{Fs}_{\mathrm{Fs}}$

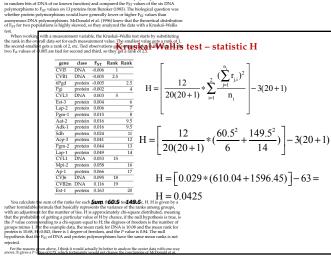






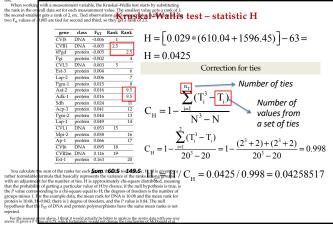


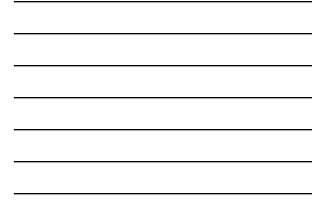
41 ic polymorphism) in two popu mald et al. (1996) collected dat



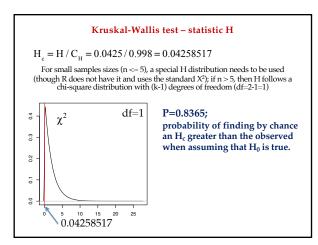


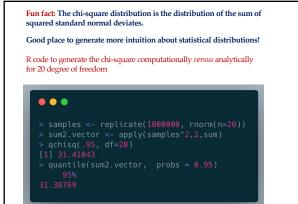
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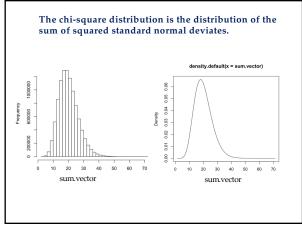




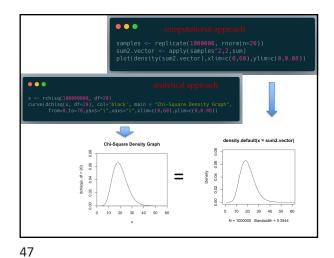
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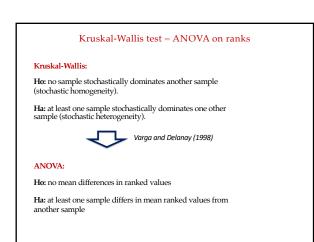
The chi-square distribution is the distribution of the sum of squared standard normal deviates.

fun fact: The F distribution is the ratio of two (scaled) chi-square distributed values. The scaling is done by appropriate division of degrees of freedom.

A general solution to rankbased tests

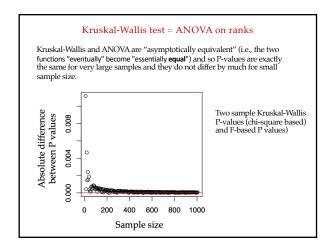
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	s equivalent (close enough) to an NOVA on ranks
Ho: no sample stochastically do (stochastic homogeneity).	ominates another sample
Ha: at least one sample stochas sample (stochastic heterogeneit	tically dominates one other ty).
of the rank sample means . The stochastic homogeneity can be	uivalent to the equality of the expected values is finding implies that the null hypothesis of e tested by an ANOVA performed on the rank ly equivalent to doing a Kruskal-Wallis H test."
of the rank sample means . The stochastic homogeneity can be	is finding implies that the null hypothesis of e tested by an ANOVA performed on the rank
of the rank sample means. The stochastic homogeneity can be transforms, which is essentiall	is finding implies that the null hypothesis of e tested by an ANOVA performed on the rank y equivalent to doing a Kruskal-Wallis H test."







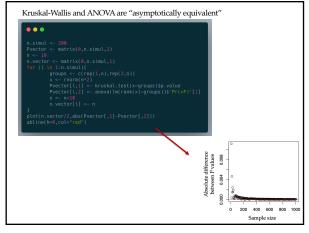


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Kruskal-Wallis and ANOVA are "asymptotically equivalent" and so P-values are the same for very large samples and they do not differ by much for small sample size. Using R code to demonstrate the asymptotic equivalence.

```
n.simul <- 200
Pvector <- matrix(0,n.simul,2)
n <- 10
n.vector <- matrix(0,n.simul,1)
for (i in 1:n.simul){
  groups <- c(rep(1,n),rep(2,n))
  x <- rnorm(n*2)
  Pvector[i,1] <- kruskal.test(x-groups)$p.value
  Pvector[i,2] <- anova(lm(rank(x)~groups))$'Pr(>F)'[1]
  n <- n + 10
  n.vector[i] <- n
}
</pre>
```

plot(n.vector/2,abs(Pvector[,1]-Pvector[,2]))
abline(h=0,col="red")





Kruskal-Wallis test = ANOVA on ranks

Kruskal-Wallis and ANOVA are "asymptotically equivalent" and so P-values are exactly the same for very large samples and they do not differ by much for small sample size.

Because of the equivalence, we can then expand nonparametric analysis based on ranks to any multi-factorial ANOVAs, regressions, MANOVA, ANCOVA, etc

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NOTE: Non-parametric tests are those that can handle non-normal data

There is a common misunderstanding in the statistical literature and among practitioners, including many biostatistics books, that non-parametric tests can also handle differences in variances among samples.

THIS IS NOT TRUE! They are also affected by variance differences among groups/treatments (i.e., homoscedasticity).

Test variance differences in ranks (almost never done in the literature)!

